Annual Veterinary Sample Kit: Collection & Shipping Instructions
Tips for Participating Veterinarians

Thank you for your participation in this groundbreaking study. We created the following list of tips based on feedback we’ve received from our Study veterinarians over the last few years. We understand the extra time and effort it takes to participate in the Study and we want to streamline the process as much as possible – you’re important to us!

If you have any other ideas, please feel free to share them with the Study team!

If you have questions or need assistance, the customer care team is a great resource. They’re available 9 a.m. — 6 p.m. ET at 855.4GR.DOGS (855.447.3647), or grdogs@caninelifetimehealth.org.

• As soon as possible after the annual Study examination and sample collection, please complete the veterinary questionnaire for your patient. (On the morrisanimalfoundation.org homepage, click on the GRLS icon in the upper right corner, and scroll down to log in). Timely questionnaire completion helps our Study team stay up-to-date on Study operations.

• We’ve found that writing your login username and password in each Study patient’s chart can make the login process quicker.

• For the annual questionnaire, you’ll need to report on all medications dispensed and vaccines given over the previous 12 months. You’ll also be reporting on any new health event. If your software system does not show problem lists, medications and vaccines separately, consider maintaining an annual list for each category within the patient chart for easy review. Remember that you’ll need the vaccine manufacturer and lot number.

• Don’t forget that anyone can enter the annual exam data - a knowledgeable staff member can input any information online from your medical records.

• Many of our Study veterinarians find that adding extra time on the day of the Study for completing their questionnaire is a good way of getting all their Study requirements done quickly.

We appreciate the time and care our Study veterinarians take to help make our study a success – we couldn’t do it without you!
Sample Collection Procedures

NOTE: The annual kit is organized into separate bags: one for clinical testing (ZNLabs) and one for the biorepository (Fisher). You have a separate bag with blood draw supplies. Cap colors for specimen transport tubes may vary due to recent shortages of medical supplies.

Blood Collection

Blood samples should be collected using the Vacutainer blood collection system included in this sample kit. For instructions on how to use the Vacutainer blood collection system, see Appendix 1.

You may use either of the following:

• The butterfly catheter attached to the Vacutainer needle holder using a cephalic vein or lateral saphenous vein

• The straight needle attached to the Vacutainer needle holder using a jugular vein.

Serum Isolation

Allow the four serum tubes (red tops) to clot for at least 45 minutes and then centrifuge per your clinic’s normal protocol (we recommend 1100–1300 RCF for 10 minutes).

After centrifugation, remove the Vacutainer cap from each of the four serum tubes (red tops) and using the 3 mL transfer pipette provided (DO NOT POUR), transfer the serum as follows:

• Place 1.5 mL of serum into the 3 mL serum tube. The black line on the left side of the tube label indicates the appropriate fill volume (white top; no barcode).

Verify the information and print the date on the label.

• Place 10 mL of serum into the 10 mL biorepository serum transport vial (screw-top; barcode on label: 094xxxxxx20yy, SERUM).

NOTE: the cap color of the serum transport tube may vary.

If additional serum is needed to achieve the required volumes, try re-centrifuging the blood remaining in the tubes. Once the appropriate amount of serum has been collected, discard the clotted red-top tubes.

Blood Collection

Blood samples should be collected using the Vacutainer blood collection system included in this sample kit. For instructions on how to use the Vacutainer blood collection system, see Appendix 1.

You may use either of the following:

• The butterfly catheter attached to the Vacutainer needle holder using a cephalic vein or lateral saphenous vein

• The straight needle attached to the Vacutainer needle holder using a jugular vein.

You will need to collect at least 56 mL of blood.

If necessary, use a needle and syringe (not supplied) to collect additional blood to top off any underfilled tubes.

Serum Isolation

Allow the four serum tubes (red tops) to clot for at least 45 minutes and then centrifuge per your clinic’s normal protocol (we recommend 1100–1300 RCF for 10 minutes).

After centrifugation, remove the Vacutainer cap from each of the four serum tubes (red tops) and using the 3 mL transfer pipette provided (DO NOT POUR), transfer the serum as follows:

• Place 1.5 mL of serum into the 3 mL serum tube. The black line on the left side of the tube label indicates the appropriate fill volume (white top; no barcode).

Verify the information and print the date on the label.

• Place 10 mL of serum into the 10 mL biorepository serum transport vial (screw-top; barcode on label: 094xxxxxx20yy, SERUM).

NOTE: the cap color of the serum transport tube may vary.

If additional serum is needed to achieve the required volumes, try re-centrifuging the blood remaining in the tubes. Once the appropriate amount of serum has been collected, discard the clotted red-top tubes.

Urine Collection

Collect a minimum of 10 mL of urine via free catch or cystocentesis. A transfer pipette is provided for your convenience to:

• Place a minimum of 5 mL of urine into the urine tube for clinical testing. This tube has an account number and no barcode on the label.

Verify the information on the label and print the date on the label.

• Place a minimum of 5 mL of urine into the biorepository urine tube (barcode: 094xxxxxx30yy, URINE).

NOTE: the cap color of the urine transport tube may vary.

If additional urine is needed to achieve the required volumes, try re-centrifuging the remaining urine in the tubes. Once the appropriate amount of urine has been collected, discard the clotted red-top tubes.

NOTE: The annual kit is organized into separate bags: one for clinical testing (ZNLabs) and one for the biorepository (Fisher). You have a separate bag with blood draw supplies. Cap colors for specimen transport tubes may vary due to recent shortages of medical supplies.

If there are complications and you are not able to obtain all the blood required in one visit, we request that all samples be redrawn at a later date using a Redraw Kit. To request a redraw kit, please call the Study team at 855.4GR.DOGS (855.447.3647).

Using the Vacutainer holder and either the butterfly or straight needle provided, draw blood in the following order:

1. Fill the 3 mL EDTA tube (purple top) and invert several times to prevent clotting.

   • Verify the information and print the date on the label.

2. Fill the 10 mL EDTA biorepository tube (purple top; barcode: 094-xxxxxx10yy, EDTA) and invert several times to prevent clotting.

3. Fill the four 10 mL serum tubes (red tops; no barcodes).

You will need to collect at least 56 mL of blood.

If necessary, use a needle and syringe (not supplied) to collect additional blood to top off any underfilled tubes.

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Sample Collection Procedures (Cont.)

Fecal Collection
If the owner did not bring the dog’s fecal sample to the visit, please obtain a sample from the patient during the visit. Transfer a marble-sized sample (approximately 1-2 g) of feces into each of the following:

- Large 30-ml fecal collection tube for clinical testing.
  - Verify the information on the label (no bar code) and print the date on the label.
- Small biorepository fecal collection tube (label contains barcode: 094xxxxxx60yy, FECAL).

Toenail Trimmings Collection
Collect 5–10 toenail trimmings from the dog.

- Place the toenail trimmings into the biorepository 2 mL plastic cryogenic tube (barcode: 094xxxxxx50yy, NAIL).

Hair Collection
Cut a lock of hair ≈1/4” in diameter (about the diameter of a wooden pencil) and a minimum of 2” long as close to the root as possible.*

- Place the lock of hair into the 3.5 mL biorepository tube (orange screw-top; barcode: 094xxxxxx40yy, HAIR).

* Please ask the owner for the preferred collection site.

Shipping Instructions

Placing A Service Call For FedEx Shipment(s)
Place a service call to FedEx directly at 800.463.3339. Let them know you have packages for pickup and tell them that the shipping cost is being billed to Morris Animal Foundation with a “Billable Stamp.” If your clinic is inadvertently charged any fees, please contact the Study team.

All samples must be collected and shipped the same day.
Sample Packaging & Shipping

Biorepository Shipment

Complete the “Shipment Inventory Form (Fisher BioServices)—Golden Retriever Lifetime Study” and include the patient information, collection date, collection times, sample shipment inventory and any optional comments.

- Place the following three samples into the Styrofoam shipping container*
  - Urine—10 mL transport tube with “Urine Only” label
  - 10 mL of EDTA blood—purple-top Vacutainer
  - 10 mL of serum—serum transport tube with “Serum Only” label

- Close the Styrofoam shipping container and place it inside the zip-closure Lab-Loc Biohazard bag. Also place the following five items inside the zip-closure Lab-Loc Biohazard bag:
  - Small fecal sample tube
  - Nails cryogenic tube
  - Hair cryogenic tube
  - Absorbent paper towel
  - Shipping inventory form

- Place the biorepository zip-lock bag inside the FedEx Clinical Pak bag that has the Fisher Biorepository mailing stamp pre-affixed.

* NOTE: All tubes for the biorepository shipment are barcoded.

Sample Packaging & Shipping (Cont.)

CLINICAL Diagnostics Shipment

Verify all four tubes (no barcodes) are properly filled and labeled, including the date. Place the large fecal, 3 mL serum (white top), 3 mL EDTA (purple top) and 5 mL urine tubes into a zip-closure biohazard bag.

- In the top header of the ZNLabs Diagnostics requisition form, provide the following information: your clinic name and address, email, phone and fax number.
- Complete the information (doctor, date, client name, pet name, species, breed and age) on the form.
- All laboratory tests are ordered with code 4761; this is a billing code for the Study, so you must use this requisition form and verify that the Study code is preprinted on the form. Failure to do so may result in charges to your clinic. No additional tests may be run with this order.

Package the samples for shipment to ZNLabs:

- Place the completed ZNLabs Diagnostics requisition form into the biohazard zip-closure bag (with the necessary samples).
- Place the zip-closure bag into the small shipping box and tape the box closed.
- Place the small shipping box in the FedEx Clinical Pak bag that has the ZNLabs mailing stamp pre-affixed.

Including the ZNLabs requisition form ensures that your clinic will receive the lab results.
Entering Examination Information on the Study Website

Annual Examination
Don’t forget! Please collect the information from your patient’s examination and log on at morrisanimalfoundation.org to complete the annual questionnaire form for your patient. Click on the GRLS icon at the top right of the homepage, and scroll down to log in.

You will receive the test results from the samples you ship to ZNLabs within three to five business days on your patient’s record through your online Study account at morrisanimalfoundation.org. The results are posted under the tab labeled “Lab Results.” The panel that is completed at this appointment includes: SuperChem, CBC, urinalysis, Total T4, heartworm antigen and fecal O&P.

Additional Study Reminders
Notify the Study team immediately if you suspect or diagnose malignancy in a Study dog. Full sample collection, including lymph node biopsy in suspected lymphoma cases, is required BEFORE treatment begins. Once malignancy is confirmed, you must complete a Malignancy Related Questionnaire. This additional form will be found by logging on to your veterinarian user account on the Study website.

If you have any questions, please do not hesitate to email the Study team at grdogs@caninelifetimehealth.org or call toll-free at 855.4GR.DOGS (855.447.3647). We are here to help you!

Appendix 1

Safety-Lok Butterfly Collection Needle
1. Check to ensure that the female luer adapter is securely attached to the male luer adapter and securely attached to the holder.
2. Remove the needle sheath and, while holding the wing, perform venipuncture according to your facility’s established procedures. Then fill the required number of blood tubes.
3. Withdraw the blood collection set by grasping the translucent yellow safety-shield grip.
4. With the opposite hand, grasp the tubing and push the yellow shield forward until the safety shield is locked in place (you will hear a click).
5. Discard both the needle and the holder (as one unit) into your sharps container.

Eclipse Standard Collection Needle
1. Screw the holder onto the needle until it fits securely and the needle is fully seated onto the holder.
3. Twist and pull the colored needle cap straight off.
4. Perform venipuncture according to your facility’s established procedures and fill the required number of blood tubes.
5. Discard both the needle and the holder (as one unit) into your sharps container.

Read all instructions before collecting samples.
About Morris Animal Foundation

Morris Animal Foundation is a nonprofit organization that invests in science to advance animal health. The Foundation is a global leader in funding scientific studies for companion animals, horses and wildlife. Since its founding in 1948, Morris Animal Foundation has invested in studies that have led to significant breakthroughs in diagnostics, treatments, preventions and cures to benefit animals worldwide. Learn more at morrisanimalfoundation.org.

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Please contact the Study team at 855.447.3647 for advice or assistance with any sample submission or to request replacement or additional supplies.